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Dietary vitamin D₃ supplementation protects laying hens against lipopolysaccharide-induced immunological stress

Yanqiang Geng, Qiugang Ma, Zhong Wang* and Yuming Guo

Abstract

Background: The effects of vitamin D on the immune function of laying hens are not well understood. This study investigated the effects of vitamin D₃ (VD₃) on laying performance and immunological functions in laying hens under *Escherichia coli* lipopolysaccharide (LPS) challenge.

Methods: In experiment one, 360 Jinghong-1 strain layers (32 weeks) were randomly divided into four groups with six replicates per group and 15 hens per replicate. Hens were fed a basal diet supplemented with different levels of VD₃ (0; 500; 1500; or 3000 IU VD₃/kg of diet) for 10 weeks to determine laying performance, egg quality, and other parameters. In experiment two, 24 Jinghong laying hens (32 weeks) were fed basal diets with either 0 or 3000 IU VD₃/kg of diet. After 10 weeks of feeding, six hens from each treatment were injected intravenously with 8 mg/kg of body weight of either LPS or saline. Blood and spleen samples were obtained for immune parameter analysis 4 h after injection.

Results: VD₃ deficiency reduced egg production and egg quality; in addition, feed intake and feed-to-egg ratio increased. No significant differences were observed in these parameters except eggshell strength between dietary VD₃ supplemental levels at 500; 1500; and 3000 IU VD₃/kg of diet. VD₃ deficiency increased serum hormone (calcitonin, parathyroid hormone, estradiol, and progesterone) and cytokine (IL-6, IL-10) levels, the ratio of IFN-γ to IL-4, myeloperoxidase activity and total IgG content in the serum, and upregulated the blood CD3⁺ T cell population. Splenic retinoid X receptor (RXR), nuclear factor-κB (NF-κB), inducible nitric oxide synthase (iNOS), and polymeric immunoglobulin receptor (pIgR) gene mRNA levels were upregulated in VD₃-deficient hens. VD₃ deficiency significantly reduced serum Follicle stimulating hormone (FSH) and Luteinizing hormone (LH) concentrations and the number of CD4⁺CD25⁺ T cells in the blood. These changes were completely normalized by VD₃ sufficiency. LPS reduced serum LH concentration, splenic lysozyme, and pIgR gene mRNA levels. LPS induced an increase in total serum IgM levels and the percentage of CD8⁺ T cells in the blood. The changes were completely reversed by VD₃ addition.

Conclusion: VD₃ supplementation could protect laying hens not only from VD₃ deficiency but also from immunological stress.

Keywords: Vitamin D₃, *Escherichia coli* lipopolysaccharide, Laying hens, Immunomodulation, Serum hormone

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Background

Vitamin D₃ (VD₃), also known as cholecalciferol, is the inactive form of vitamin D that can be ingested through dietary intake or be generated endogenously in the skin of animals exposed to UV light. VD₃ is converted into its active form, 1, 25-dihydroxycholecalciferol (1,25-(OH)₂D₃), following a two-step hydroxylation process mediated by two key enzymes, 25-hydroxylase and 1 α -hydroxylase. The first hydroxylation step occurs in the liver by 25-hydroxylase, which hydroxylates cholecalciferol at the 25-C position to form 25-hydroxycholecalciferol (25(OH)D). The subsequent hydroxylation of 25-hydroxycholecalciferol occurs at the 1-C position and is mediated by 1 α -hydroxylase in the kidneys to produce 1,25-(OH)₂D₃, the active form of vitamin D [1, 2]. This active form of vitamin D is thought to act via binding to the nuclear vitamin D receptor protein (VDR). The VDR then binds to target DNA sequences as a heterodimer with the retinoid X receptor (RXR), recruiting a series of coactivators, inducing target gene expression [3, 4]. Vitamin D is an essential nutrient and plays an integral role in maintaining calcium and phosphorus homeostasis, skeletal health, and muscle development [5–7]. Interestingly, increasing evidence derived from research in murine and human systems has demonstrated that vitamin D plays an important role in maintaining normal immune function and crosstalk between the innate and adaptive immune systems by influencing vitamin D receptors and activating enzymes [8–10]. Furthermore, VD₃ has been shown to possess immunomodulatory, antioxidant, anti-inflammatory, antibacterial, antiviral, anti-allergy and cancer prevention properties [11–13]. In mammals, VD₃ insufficiency and deficiency result in not only dysfunction of the innate and adaptive immune systems but also promotes micro-inflammation, as well as an increased risk of viral or bacterial infections [14–17]. Clinical trials of inexpensive VD₃ supplements at appropriate doses show the vitamin's effectiveness in the prevention and control of the diseases and inflammation through enhancing innate immunity, inducing antimicrobial peptide synthesis and nitric oxide production, dampening excessive inflammation responses, or decreasing the production of pro-inflammatory cytokines. The vitamin acts directly on T lymphocytes or antigen presenting cells [7, 8, 18–20]. In birds, the role of VD₃ in calcium and phosphorus metabolism is crucial for its well-documented involvement in bone development and eggshell formation in laying hens. However, the immunomodulatory, anti-inflammatory and anti-coccidia roles of VD₃ or its metabolites have also been demonstrated in chickens [21–24] and chicken cells [23, 25]. In addition, Rodriguez-Lecompte et al. [26] reported that vitamin D or 25(OH)D, both have a robust immunomodulatory property with a more favorable helper T cell (Th2) response in broiler chickens. Aslam et al. (1998) [27] observed that vitamin D deficiency depresses the cellular immune responses in

young broiler chicks. Therefore, research into the immunomodulatory, anti-inflammatory, and anti-infection roles of vitamin D continues to highlight novel opportunities for the promotion of animal and poultry health based on implementation of vitamin D supplemented nutritional regimens [28].

Laying hens reared under intensive commercial conditions are often threatened by the large amounts of pathogen-associated molecular patterns, such as lipopolysaccharides (LPS) derived from gram-negative bacteria and lipoteichoic acid derived from gram-positive bacteria. Inhalation of environmental gram-negative bacteria (their endotoxins in particular) has been suggested to be a major poultry health problem. Administration of LPS to chickens stimulates the immune system, activating the transcription of nuclear factor (NF)- κ B, releasing acute phase proteins and proinflammatory cytokines, causing the expression of toll-like receptors 2 (TLR2) and 4 (TLR4), and modulation of antibody responses [29], besides causing clinical symptoms such as fever, anorexia, and decreased growth [29, 30].

Interest in using nutritional strategies to modulate the immune functions caused by LPS in chickens has increased in the last decade. Significant accumulated evidence from mammalian studies have demonstrated that supplementation with VD₃ inhibits inflammatory cytokine production in LPS-induced acute lung injury [31] and alleviates LPS-induced renal oxidative stress by regulating oxidant and antioxidant enzyme genes [12]. Some reports have shown that VD₃ or 25-hydroxycholecalciferol exhibit anti-inflammatory effects in broilers or chicken immune cells following LPS administration [21–23]. However, little information has been reported about the effects of dietary VD₃ supplementation or deficiency on immune responses and serum biochemical indexes of laying hens challenged with LPS or the underlying mechanisms of action. Thus, our study was designed to investigate the effects of dietary VD₃ supplementation and deficiency on egg performance, blood T lymphocyte subset percentages, serum biochemical constituents, serum natural antibodies and cytokines, and mRNA abundance of splenic immune-related genes in laying hens (32 to 42 weeks of age) subjected to acute *Escherichia coli* LPS challenge, and to explore how laying hens respond to those manipulations. Such investigations may provide evidence of whether dietary VD₃ supplementation exerts dual roles in laying hens following immune challenge, by improving egg performance and the immune function.

Methods

Animal ethics statement

This study, which complied with institutional and national guidelines for the care and use of animals, was approved by the Committee of Animal Experiments of

China Agricultural University. All efforts were made to minimize animal suffering.

Animal model and experimental treatments

Two experiments were conducted separately in this study. Experiment one was designed to study the effects of dietary VD₃ levels on egg production, egg quality, and serum 25(OH)D levels of laying hens. A total of 360 Jinghong laying hens (Beijing Yukou Poultry Co., Ltd., China) at 32 weeks of age with similar weight, production performance, and genetic background were randomly assigned to four groups with six replicates per group and 15 birds per replicate. Hens were fed a corn–soybean meal basal diet supplemented with different levels of VD₃ (0; 500; 1500; or 3000 IU VD₃/kg) of diet. The basal diet was formulated to satisfy the Chinese Feeding Standard of Chickens (NY/T33–2004) and nutrient requirements of laying hens consuming 110 g of feed per day. The basal composition of the diet is shown in Table 1. The VD₃ requirement of laying hens according to the Chinese Feeding Standard of Chickens (2004) is 1600 IU/kg of diet; however, the amount of VD₃ added for laying hens in practical production is 3000 IU/kg of diet. The analyzed VD₃ content of the laying hen basal diet was 0.00 IU/kg of diet (from natural VD₃ in feed ingredients); no synthetic VD₃ was included in the basal diet. Hens were housed in an experimental layer farm and kept in three-layer complete ladder cages at three birds per cage (40 × 45 × 45 cm) equipped with water nipples.

Table 1 Composition and nutrient levels of the experimental vitamin D₃-deficient diets

Ingredients (%)	Composition	Calculated nutrient levels ^c	Value
Corn (7.8% crude protein)	66.45	Metabolic energy(MJ /kg)	11.30
Soybean meal (43% crude protein)	22.80	Crude Protein(%)	15.52
Limestone	8.20	Calcium(%)	3.60
calcium hydrophosphate	1.70	Available phosphorus(%)	0.39
Sodium chloride	0.30	Lysine(%)	0.75
DL-methionine (98%)	0.12	Methionine(%)	0.37
Choline chloride (50%)	0.10	Met+Cys(%)	0.68
Vitamin premix ^a	0.03	Tryptophane(%)	0.18
Trace elements premix ^b	0.30	Threonine(%)	0.57

^aProvided per kilogram of diet: vitamin A, 6000 IU; vitamin E, 21 IU; vitamin E, 21 IU; vitamin K3, 4.2 mg; vitamin B1, 3 mg; vitamin B2, 10.2 mg; folic acid, 0.9 mg; pantothenic acid, 15 mg; niacin, 45 mg; vitamin B6, 5.4 mg; vitamin B12, 24 µg; biotin, 0.15 mg. VD₃ added alone. Prepared with a small mixing machine after premix mixing. High performance liquid chromatography to detect the actual content, and the calculation of the value of the match before the feed preparation

^bMineral premix provided per kilogram of complete diet: copper, 6.8 mg; iron, 66 mg; zinc, 83 mg; manganese, 80 mg; iodine, 1 mg; Se 0.3 mg

^c Nutrient levels were calculated from data provided by Feed Database in China (2012)

During the test period, the birds were fed three times each day and supplied with water ad libitum. Additionally, a lighting procedure of 16 h illumination and 8 h darkness was maintained throughout the experiment. The average temperature in the chicken house was 24.5 ± 0.5 °C. Feed consumption was recorded weekly throughout the entire experimental period. Egg number, total egg weight and broken eggs of each replicate were recorded daily and calculated as percentage hen-day egg production, average egg weight, daily egg mass. and broken egg rate. Egg mass was calculated as egg weight × egg production. Feed efficiency (feed conversion rate, FCR) was calculated as grams of feed consumed per gram of egg mass (egg production/100 × egg weight) produced. Mortalities and health status were visually observed and recorded daily throughout the entire experimental period.

Experiment two was designed to study the effects of VD₃ supplementation on immune response and serum reproductive hormones in laying hens challenged with LPS. After 10 weeks of a feeding trial, 12 laying hens from the control (without VD₃ supplementation) and 12 laying hens from the VD₃-supplemented group (supplemented 3000 IU VD₃/kg of diet) were randomly selected, and six hens from each dietary treatment were injected intravenously with either 8.0 mg/kg body weight of *E. coli* LPS (serotype O111:B4, Sigma-Aldrich Inc., St. Louis, MO, USA) diluted in sterile saline. Feed was removed from the birds after injection. Four hours after LPS or saline injection, blood samples were collected via wing venipuncture and divided into two aliquots (2 to 3 mL each); one 4-mL serum vacutainer tube and one 4-mL vacutainer tube coated with K₂EDTA (BD BioSciences, San Jose, CA, USA). These blood samples were kept on ice during collection, whereas the blood samples for serum were clotted at room temperature during collection, centrifuged at 12,000×g for 5 min at 4 °C, and serum was recovered for biochemistry analysis. After the blood collection, all birds were killed by cervical dislocation and splenic samples were collected and frozen immediately with liquid nitrogen and stored at – 80 °C until further analysis.

Flow cytometry analysis

The percentages of CD3⁺, CD4⁺, CD8⁺, and CD4⁺CD25⁺ cells among peripheral blood lymphocytes were analyzed by flow cytometry as described previously [32]. Briefly, the blood from K₂EDTA vacutainer tubes was diluted 1:1 with 1 × PBS and held on ice. The blood suspensions were carefully layered into centrifuge tubes containing an equal volume of Ficoll to form a discrete layer above the Ficoll. The tubes were centrifuged at 220×g for 30 min at room temperature, and the mononuclear layers (buffy coats) were removed, transferred to different centrifuge tubes, and washed twice in 1 × PBS. Immediately, the cells were counted on a hemocytometer

using the trypan blue exclusion assay (Sigma-Aldrich Inc. USA) and the cell suspensions were adjusted to 1×10^6 viable cells per mL of $1 \times$ PBS. The following monoclonal antibodies were used in immunofluorescence assays: mouse anti-chicken-CD3-SPRD-labeled, mouse anti-chicken-CD4-FITC-labeled, mouse anti-chicken-CD8-RPE-labeled (Southern Biotechnology Associates Inc., Birmingham, AL, USA) and mouse anti-chicken CD25-Alexa Fluoro 647-labeled (AbD Serotec, USA). Negative isotype control IgG (mouse IgG1 κ -SPRD, mouse IgG1 κ -FITC, mouse IgG1 κ -R-PE, and HuCAL Fab-dHLX-MH negative control-Alexa Fluoro 647-labeled (AbD Serotec, USA) were diluted in PBS (pH 7.2). Using 96-well round-bottom plates, 100 μ L of each cell suspension was plated in duplicate. Each of the fluorescein-labeled antibodies was added to the respective wells, and the stained cells were incubated for 30 min at 4 °C in the dark. The cells were washed twice in cold $1 \times$ PBS and centrifuged for 30 min at 1800 g to remove any traces of unbound antibodies. They were then transferred to 5-mL polystyrene round-bottom tubes for analysis. A total of 10,000 cells per sample were conducted using a Coulter XL (Beckman Coulter Corp., Fullerton, CA, USA) at Xi-Yuan Traditional Chinese Medicine Hospital, Chinese Academy of Medicine Science, China. The percentages of CD3⁺ T, CD4⁺ T, CD8⁺ T and CD4⁺ CD25⁺ T cells in PBMC were subsequently calculated.

Serum calcium and 25-hydroxycholecalciferol analysis

Serum total calcium content (mM) was measured with the UV-2000 visible spectrophotometer (Unico Instruments Co. Ltd., Shanghai, China) using commercial colorimetric assay kits (Nanjing Jiancheng Bioengineering Institute, Nanjing, China).

To determine serum 25-hydroxycholecalciferol (25(OH)D), serum samples were separated by centrifugation at 894 \times g for 15 min and frozen at -20 °C. Ultra-high-performance liquid chromatography–tandem mass spectrometry was used to assess 25(OH)D as previously described [33]. A standard curve was obtained using dilutions of a 25(OH)D standard (Iso Sciences, USA).

Serum hormone analysis

Corticosterone (CORT), calcitonin (CT), parathyroid hormone (PTH), estradiol (E2), follicle stimulating hormone (FSH), luteinizing hormone (LH), testosterone (T), and progesterone (PG) levels in the serum were determined with commercial radioimmunoassay (RIA) kits in accordance with the manufacturer's instructions (Beijing North Institute of Biological Technology, Beijing, China).

Serum complement and cytokine analysis

Serum myeloperoxidase (MPO), complement components (C3, C4, C5) were measured using a commercial kit (Nanjing Jiancheng Bioengineering Institute). Serum

concentrations of interleukins (IL-1 β , IL-2, IL-4, IL-6, IL-10), interferon- γ (IFN- γ), and tumor necrosis factor- α (TNF- α) were determined with commercially available chicken cytokine ELISA kits (Beijing North Institute of Biological Technology), according to the manufacturer's protocol. In each assay, a control recombinant chicken cytokine was diluted over the recommended detection range to generate a standard curve, and the linearity was calculated using Microsoft Excel 2013 (Microsoft Corporation, Redmond, WA, USA) to be $R^2 = 0.99$. Sample concentrations were interpolated from the standard curve.

Serum total IgG and IgM determination

Serum levels of total IgG and IgM were quantified with chicken IgG and IgM ELISA kits, respectively (Bethyl Laboratories, Inc., Montgomery, TX, USA) following the manufacturer's procedure. The serum samples were diluted 1:125,000 for IgG determination or 1:10,000 for IgM determination. The plates were read via an ELISA plate reader (SeptraMax[®] i3x Platform, Molecular Devices, LLC, San Jose, CA, USA) at 450 nm, and serum antibody concentrations were calculated using Gen 5 software (BioTek Instruments Inc., Winooski, VT, USA).

Quantitative real-time PCR for measuring immune-related gene transcript levels in the spleen

Total RNA was isolated from snap-frozen spleen tissue samples (50 mg) based on the RNeasy mini kit following the animal tissue protocol (Qiagen Sciences, Inc., Germantown, MD, USA). The purity and concentration of the total RNA were measured in a NanoDrop-2000 spectrophotometer (ThermoFisher Scientific Co., Waltham, MA, USA) using the 260:280 nm absorbance ratio. First-strand cDNA was synthesized from 2 μ g of total RNA using a PrimeScript RT reagent kit with gDNA eraser (Perfect Real Time; Takara Biomedical Technology (Beijing) Co. Ltd., Beijing, China) according to the manufacturer's instructions and stored at -20 °C until further processing. Primer sequences for chicken β -actin, toll-like receptor (TLR)-4, TLR-2, TNFSF15, IL-1 β , IL-6, IL-8, nuclear factor κ B (NF- κ B), polymeric immunoglobulin receptor (pIgR), VDR, RXR, interferon gamma (IFN- γ), lysozyme (LYZ), and inducible nitric oxide synthase (iNOS) (Table 2) were designed based upon sequences available from public databases using Primer Express, version 5.0 (Applied Biosystems, Foster City, CA, USA) and synthesized by Sangon Biotech (Shanghai) Co., Ltd., Shanghai, China. Primers were designed to span an intron to avoid genomic DNA amplification. Quantitative real-time PCR was performed using the 7500 Fast Real-Time PCR system (Applied Biosystems) and SYBR Premix Ex Taq kit (Takara Biotechnology Co. Ltd.). Reactions were conducted in a 20- μ L reaction mixture containing

Table 2 Sequences of primers for quantitative real-time PCR

Target gene	Primer sequence 5' → 3'	Product size (bp)	Annealing temperature (°C)	GenBank No.	Efficiency (%)
VDR	F: TGGGAAAGCGCATGCTGATG R: GATGCGAGACATGCAGATG	169 bp	59.0	NM_205098.1	86.5
RXR	F: GATGCGAGACATGCAGATG R: CGGGGTATTTGTGCTTG	161 bp	55.0	XM_015279790.1	96.3
TLR2	F: ACCTTCTGCACTCTGCCATT R: TGTGAATGAAGCACCCGTAA	131 bp	58.5	NM_204278.1	94.4
TLR4	F: CCACTATTCGGTTGGTGGAC R: ACAGCTTCTCAGCAGGCAAT	86 bp	59.0	NM_001030693.1	98.1
NF-κB	F: ACCCCTCAATGTGCCAATG R: TCAGCCCAGAAACGAACCTC	274 bp	59.3	NM_205129.1	80.5
TNF-α	F: CCCCTACCCTGTCCCACAA R: TGAGTACTGCGGAGGGTTCAT	67 bp	60.7	NM204267.1	107.9
IFN-γ	F: AAAGCCGCACATCAAACACA R: GCCATCAGGAAGGTTGTTTTTC	64 bp	58.8	NM_205149.1	90.4
IL-1β	F: CAGCAGCCTCAGCGAAGAG R: CTGTGGTGTGCTCAGAATCCA	86 bp	60.5	NM_204524.1	98.7
IL-6	F: AGATGGTGATAAATCCTGATGA R: CGGTTTTCTCCATAAATGAAGT	150 bp	54.5	NM_204628.1	130.4
IL-8	F: GGCTTGCTAGGGAAATGA R: AGCTGACTCTGACTAGGAACTGT	200 bp	59.0	NM_205498.1	88.6
LYZ	F: GACGATGTGAGCTGGCAG R: GGATGTTGCACAGGTTCC	225 bp	57.1	NM_205281.1	98.4
iNOS	F: GAACAGCCAGCTCATCCGATA R: CCCAAGCTCAATGCACAACCT	103 bp	59.8	NM_204961.1	78.1
pIgR	F: ATGAAGCAGAGCCAGGAGAC R: GAGTAGGCGAGGTCAGCATC	103 bp	59.7	NM_001044644.1	106.4
β-actin	F: GAGAAATTGTGCGTGACATCA R: CCTGAACCTCTCATTGCCA	282 bp	58.0	NM_205518	97.8

IL interleukin; *iNOS* inducible nitric oxide synthase; *pIgR* polyglobulin receptor; *TLR* toll receptor; *TNF-α* tumor necrosis factor; *LYZ* lysozyme; *RXR* retinoid X receptor; *VDR* vitamin D receptor

10.0 μL of SYBR Premix Ex Taq (2×) mix, 2.0 μL of cDNA, 0.5 μL of each primer (10 mmol/L), and 7.0 μL of sterile nuclease-free water. For PCR, samples were subjected to an initial denaturation phase at 95 °C for 5 min, followed by 40 cycles of denaturation at 95 °C for 30 s, and annealing and extension at 60 °C for 30 s. Melt-curve analysis was performed to confirm PCR amplification specificity. All tissue samples used in cDNA synthesis and in the following PCR amplifications were analyzed in triplicate. Gene expression levels of TLR4, TNFSF15, IL-1β, IL-6, IL-8, IFN-γ, LYZ, and iNOS were analyzed with β-actin (β-actin values were designated the VD3 sufficiency hens without LPS injection) as an endogenous control. The average gene expression of each sample relative to that of β-actin was calculated using the $2^{-\Delta\Delta Ct}$ method.

Statistical analysis

In experiment one, a replicate was used as the experimental unit. Data were subjected to one-way ANOVA using the GLM procedure of SPSS 17.0 for Windows (SPSS Inc., Chicago, IL, USA). The treatment means were separated by Duncan multiple range tests at $P < 0.05$ significance levels.

In experiment two, a completely randomized design with two dietary treatments and two levels of immunological challenge in a 2 × 2 factorial arrangement was used. A two-way ANOVA (the GLM procedure of SPSS 19.0 for Windows (IBM Corp., Armonk, NY, USA) was used to examine the interactive and main effects of LPS and dietary VD₃ levels on the dependent variables. Interactions were removed when the observed P value for interaction was above 0.10. When interactions were

significant ($P < 0.05$), differences between means were determined using Tukey's procedure. Differences with an α level of $P < 0.05$ was considered to be statistically significant.

Results

Effects of VD_3 deficiency and VD_3 sufficiency on egg production and egg quality

The results of the analysis of the production characteristics data are shown in Table 3; hens fed diets without VD_3 -supplementation showed a significant reduction ($P < 0.05$) in egg production, daily egg mass and average egg weight. There was a significant increase in daily feed intake, broken egg rates and feed conversion ratio (FCR), compared with the other three different levels of VD_3 -supplemented treatments. While statistical analysis showed that no significant ($P > 0.05$) differences in the above parameters of laying hens between dietary VD_3 supplemental levels at 500 IU; 1500 IU and 3000 IU/kg of diet.

Compared with VD_3 the deficiency group, dietary VD_3 supplementation significantly ($P < 0.05$) increased eggshell thickness and egg strength ($P < 0.01$), whereas it had no remarkable ($P > 0.05$) effect on Haugh unit, eggshell color, and yolk color (Table 4). Egg strength in laying hens with 1500 IU and 3000 IU/kg dietary VD_3 was significantly greater than that of hens fed 500 IU VD_3 /kg of diet.

Serum calcium and 25-hydroxycholecalciferol levels

There was a significant reduction in the levels of serum calcium (2.00 ± 0.98 mg/dL vs. 2.28 ± 1.17 mg/dL; $P < 0.05$; Fig. 1) in the VD_3 -deficient group compared with the VD_3 -sufficient group. Furthermore, serum 25(OH)D levels were significantly lower in VD_3 -deficient birds compared with VD_3 -sufficient birds (7.60 ± 2.85 ng/mL vs. 27.39 ± 2.97 ng/mL; $P < 0.05$; Fig. 2).

Serum hormone concentrations

The concentrations of hormones in serum showed (Table 5) that no significant diet \times challenge interactions

were found in the levels of serum CORT, CT, PTH, FSH, E2, T, and PG: LH was the exception. LPS-injected hens fed with the VD_3 -supplemented diets showed the highest levels of serum LH ($P < 0.05$) at 4 h after treatment with LPS compared with the other treated hens.

Relative to saline-injected hens, injection with LPS significantly reduced ($P < 0.05$) the concentrations of CT, E2, LH, T, and PG in the serum and significantly increased ($P < 0.05$) serum CORT concentration, whereas LPS had no significant effect ($P > 0.05$) on PTH and FSH. Compared with the VD_3 -deficient group, dietary VD_3 supplementation significantly reduced ($P < 0.05$) the concentration of serum CT, PTH, E2, and PG, and significantly increased ($P < 0.05$) serum FSH and LH content, whereas LPS had no significant influence ($P > 0.05$) on serum CORT and T levels.

Serum MPO activity

As shown in Table 6, LPS challenge resulted in lower MPO ($P < 0.05$) activity compared with unchallenged birds. However, relative to VD_3 sufficiency, VD_3 deficiency significantly ($P < 0.05$) increased serum MPO activity. Furthermore, there was a significant diet \times LPS injection interaction for serum MPO activity and IgM concentration. LPS-injected hens fed with a VD_3 -supplemented diet showed a significant reduction ($P < 0.05$) in MPO activity compared with saline-injected hens without VD_3 supplementation, whereas there were no significant differences ($P > 0.05$) compared with the other treatments.

Serum complement, IgG, and IgM concentrations

As shown in Table 6, LPS injection significantly decreased ($P < 0.05$) serum C5 activity, and total IgG and IgM concentration, compared with the saline-injected control. Total IgG was lower ($P < 0.05$) in the VD_3 -sufficient hens compared with the nonsupplemented hens. Furthermore, there was an interaction effect for total IgM concentration between LPS injection and VD_3 treatment. Total IgM at 4 h after treatment with LPS was found

Table 3 Effect of different levels of vitamin D_3 on laying performance of laying hens*

Items	Dietary VD_3 levels, IU/kg				SEM	P-value
	0	500	1500	3000		
Hen-day egg production (%)	27.95 ^b	90.30 ^a	94.03 ^a	92.29 ^a	5.93	< 0.001
Average egg weight(g)	58.39 ^b	60.43 ^a	60.44 ^a	59.93 ^a	0.26	0.007
Day feed intake (g /d /hen)	137.60 ^a	113.76 ^b	116.10 ^b	113.88 ^b	2.20	< 0.001
FCR (g feed / g egg)	9.56 ^a	2.12 ^b	2.07 ^b	2.09 ^b	0.74	< 0.001
Daily egg mass(g/hens/d)	16.30 ^b	54.55 ^a	56.83 ^a	55.31 ^a	3.62	< 0.001
Broken egg ratio (%)	5.48 ^a	0.29 ^b	0.17 ^b	0.27 ^b	0.01	< 0.001
Death ratio (%)	8.47	3.36	5.62	3.06	0.01	0.063

*Data are presented as mean \pm SEM ($n = 90$ hens /group)

^{a-c}Means within a row without a common superscript differ significantly ($P < 0.05$)

FCR g of feed / g of egg mass

Table 4 Effects of different levels of vitamin D₃ on egg quality of laying hens*

Items	Dietary VD ₃ levels, IU/kg				SEM	P-value
	0	500	1500	3000		
Eggshell thickness(mm)	0.35 ^b	0.43 ^a	0.44 ^a	0.45 ^a	0.01	< 0.001
Eggshell strength(kg/cm ²)	1.89 ^c	3.23 ^b	3.66 ^a	3.94 ^a	0.13	< 0.001
Haugh unit	80.36	80.33	82.35	80.08	0.83	0.764
Eggshell color	9.43	9.79	10.00	10.07	0.14	0.386
Yolk color	6.71	6.36	7.00	6.14	0.20	0.444

*Data are presented as mean ± SEM (n = 90 hens /group)

^{a-c}Means within a row without a common superscript differ significantly ($P < 0.05$)

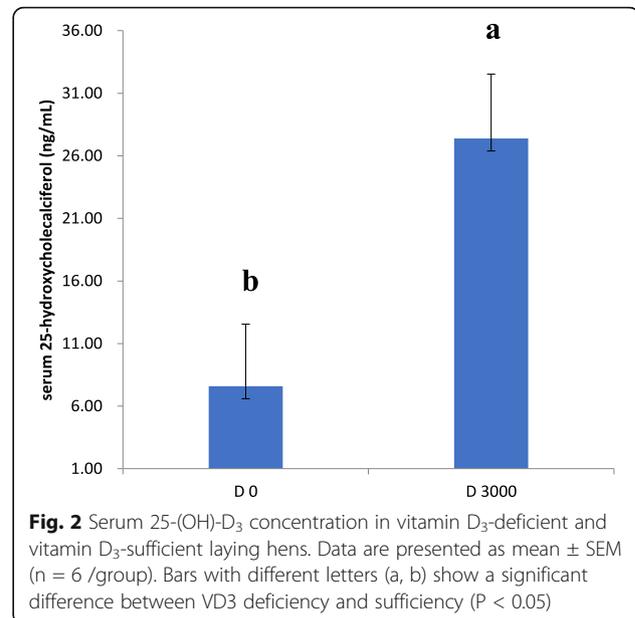
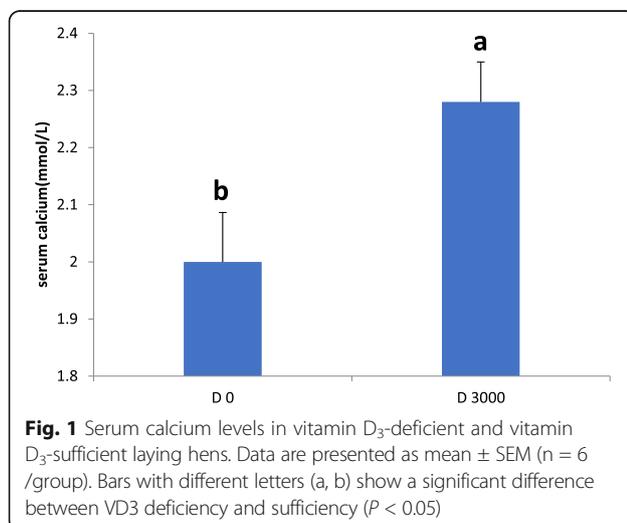
to be lower ($P < 0.05$) in VD₃-sufficient hens that were injected with LPS compared with the VD₃-added hens that were not treated with LPS, whereas there was no significant difference ($P > 0.05$) compared with the VD₃-deficient groups.

T lymphocyte population in the blood

As shown in Table 7, compared to saline-injected hens, injection with LPS significantly reduced ($P < 0.05$) the levels of CD3⁺, CD4⁺ and CD8⁺ T cells in the blood, and significantly increased ($P < 0.05$) the percentage of CD4⁺CD25⁺ T cells following LPS challenge.

Sufficient supplementation with VD₃ significantly decreased ($P < 0.05$) the percentage of CD3⁺ T lymphocytes and increased the population of CD4⁺CD25⁺ T cells ($P < 0.05$) in peripheral blood, compared with the nonsupplemented hens.

There was a significant interaction effect for the percentage of CD8⁺ T cells in the blood between dietary VD₃ treatment and LPS challenge. LPS-injected hens fed diets with supplemental VD₃ exhibited the lowest ($P < 0.05$)



percentage of CD8⁺ T cells in the blood compared with the other three treatment groups.

Serum cytokine levels

To determine whether VD₃ could exert an anti-inflammatory effect in hens, we assessed the effect of VD₃ on LPS-induced inflammatory response by measuring TNF- α , IFN- γ , IL-1 β , IL-2, IL-4, IL-6, IL-8, and IL-10 concentrations in serum treated with LPS with or without VD₃. Compared with the saline-injected control hens, LPS injection significantly ($P < 0.05$) increased the levels of TNF- α , IL-1 β , IL-2, IL-4, IL-6 and IL-8 in the serum (Table 8). Supplementation of VD₃ significantly ($P < 0.05$) reduced serum IL-6 and IL-10 amounts and the ratio of IFN- γ to IL-4 compared with the VD₃-deficient grouping.

Splenic immune-related gene expression

The effects of dietary supplementation with VD₃ on VDR, RXR, TLR-2, TLR-4, NF- κ B, TNF- α , IFN- γ , IL-1 β , IL-6, IL-8, LYZ, iNOS, and pIgR gene transcript abundance in the spleen of hens after LPS exposure are presented in Table 9. The expression levels of the TLR4, NF- κ B, and IL-6 genes were remarkably increased ($P < 0.05$) by LPS exposure as compared with the unexposed control group. In the absence of LPS, the addition of VD₃ remarkably ($P < 0.05$) prevented the upregulation of RXR- α , LYZ, iNOS, and pIgR genes expression in the spleen compared with VD₃-deficient hens, while in the presence of LPS, supplemental VD₃ upregulated ($P < 0.05$) splenic LYZ and pIgR gene mRNA levels but down-regulated ($P < 0.05$) the expressions of TLR4 and NF- κ B gene in the spleen

Table 5 Effect of vitamin D₃ on serum biochemical and hormones of laying hens infected with LPS*

Items	Dietary VD ₃ levels, IU/kg				P-values			
	0		3000		SEM	VD ₃	LPS	Interaction
	Saline	LPS	Saline	LPS				
CORT (nmol/L)	78.79	91.37	76.78	82.78	5.00	0.169	0.036	0.295
CT (pg/ml)	222.50	191.98	182.17	171.34	5.61	0.001	0.016	0.216
PTH (ng/dl)	78.47	75.94	56.28	45.04	3.89	< 0.001	0.133	0.331
FSH (mIU/ml)	9.27	6.85	13.10	13.90	0.83	< 0.001	0.344	0.075
LH (pg/ml)	9.04 ^{bc}	7.13 ^c	10.36 ^b	15.46 ^a	0.82	< 0.001	0.041	< 0.001
E2 (pg/ml)	126.41	33.81	81.81	21.00	11.01	0.035	< 0.001	0.219
P (pg/ml)	2.36	1.32	1.86	0.69	0.17	0.002	< 0.001	0.693
T (ng/dl)	71.42	33.73	86.19	28.65	6.27	0.423	< 0.001	0.112

*Data are presented as mean ± SEM (n = 6 hens /group)

^{a-c}Data in the same row with no common superscript differ significantly (P < 0.05)

SEM standard error of the mean; LPS lipopolysaccharide; CORT cortisol; CT Calcitonin; PTH parathyroid hormone; FSH Follicle stimulating hormone; E2 Estradiol; LH Luteinizing hormone; T testosterone; P progesterone

relative to the VD₃-sufficient hens. There was a significant diet × challenge interaction for TLR2, pIgR, and LYZ expression in the spleen. LPS challenge significantly down-regulated (P < 0.05) splenic TLR2, LYZ, and pIgR gene mRNA levels, and markedly up-regulated (P < 0.05) NF-κB gene mRNA levels, while these changes were normalized by VD₃ supplementation. However, feeding with VD₃ did not alter (P > 0.05) the expression levels of VDR, TLR-4, TNF-α, IL-1β, IL-6, and IL-8 in the spleen of hens compared with the nonsupplemented hens.

Discussion

Our study showed that VD₃ deficiency significantly reduced laying performance and egg quality. However, the addition of VD₃ at 500 IU; 1500 IU; and 3000 IU/kg to the basal diet has been reported to significantly increased egg production, egg weight, and egg quality; consequently, FCR was improved compared with the VD₃-deficient hens. Moreover, hens fed diets containing 1500 IU and 3000 IU VD₃/kg of diet had stronger

eggshells than birds given 500 IU VD₃/kg of diet. Consistent with our findings, similar findings on production performance were reported in previous studies [34–36], where production rate, daily egg weight, and feed intake did not differ. Other studies [37, 38] have reported no significant differences in laying performance and egg quality among different level of VD₃ supplementation in the diet. Thus, our results indicated that suitable supplemental dosage of VD₃ for middle-phase laying hens is 500 to 3000 IU/kg of diet. Additionally, decreased egg production and egg weight as well as higher mortality rates in the VD₃-deficient hens were probably because of changes in intestinal microbiota, influences on liver lipid metabolism, and reduced energy utilization efficiency (unpublished data).

25(OH)D, is the major circulating form of vitamin D and is used as indicator of vitamin D status [39]. In our study, the concentration of 25(OH)D and calcium in serum were significantly lower in VD₃-deficient birds compared with VD₃-sufficient birds, which was in agreement with previous studies using mouse models [40].

Table 6 Effect of vitamin D₃ on serum MPO, complement and natural antibody of laying hens infected with LPS*

Items	Dietary VD ₃ levels, IU/kg				P-values			
	0		3000		SEM	VD ₃	LPS	Interaction
	Saline	LPS	Saline	LPS				
MPO (ng/ml)	39.23 ^a	21.04 ^b	24.29 ^b	22.62 ^b	1.98	0.011	< 0.001	0.003
C3 (g/L)	0.87	0.78	0.88	0.76	0.03	0.896	0.054	0.751
C4 (g/L)	0.37	0.23	0.25	0.20	0.03	0.141	0.058	0.367
C5 (μg/ mL)	73.94	55.7	79.06	52.69	4.47	0.896	0.012	0.615
IgG (mg/mL)	21.4	17.96	18.76	15.25	1.17	0.038	0.009	0.977
IgM (μg/mL)	253.01 ^b	252.67 ^b	328.70 ^a	163.04 ^c	35.63	0.851	0.041	0.043

*Data are presented as mean ± SEM (n = 6 hens /group)

^{a-c}Data in the same row with no common superscript differ significantly (P < 0.05)

SEM standard error of the mean; LPS lipopolysaccharide; MPO myeloperoxidase; C3 complement 3; C4 complement 4; C5 complement 5; IgG immunoglobulins G; IgM immunoglobulins M

Table 7 Effect of vitamin D₃ on peripheral blood mononuclear cell of laying hens infected with LPS*

Items	Dietary VD ₃ levels, IU/kg				P-values			
	0		3000		SEM	VD ₃	LPS	Interaction
	Saline	LPS	Saline	LPS				
CD3	73.68	37.02	63.30	23.02	4.96	0.042	< 0.001	0.750
CD4	55.17	27.13	46.13	17.73	3.91	0.087	< 0.001	0.972
CD8	9.80 ^a	5.90 ^b	10.20 ^a	2.23 ^c	0.79	0.080	< 0.001	0.033
CD4 ⁺ CD25 ⁺	37.70	68.60	47.83	80.68	4.23	0.039	< 0.001	0.848
CD4 ⁺ /CD8 ⁺	5.98	6.06	4.62	12.62	1.19	0.236	0.072	0.077

*Data are presented as mean ± SEM (n = 6 hens /group)

^{a-c} Data in the same row with no common superscript differ significantly (P < 0.05)

SEM standard error of the mean; LPS lipopolysaccharide

Conversely, the concentrations of 25(OH)D increased in response to dietary VD₃ supplementation in our study, were accompanied by increases in serum concentrations of calcium, as shown by other researchers' findings in chickens [41–43]. The role of VD₃ in calcium and phosphorus metabolism is crucial for its well-documented involvement in bone and eggshell formation in layer hens [44]. Our results have shown that hens fed diets without VD₃ supplementation showed a significant reduction in egg production as well as poor egg quality, whereas supplementation with VD₃ significantly improved egg production and eggshell thickness and strength. These changes were accompanied by increased serum calcium and 25(OH)D concentration, implying that VD₃ deficiency might affect calcium and VD₃ metabolism, and disturb the process of eggshell formation.

This study further investigated the effects of VD₃ deficiency and sufficiency on blood hormones and immune function in laying hens challenged with *E. coli* LPS. CORT, as a glucocorticoid, has a potential immune-suppressive and pro-inflammatory cytokine regulatory effect in animals and poultry [45]. Hence, changes in plasma or serum CORT concentration have been commonly used to assess

stress and poultry welfare. However, follicle development and ovulation is controlled by complex endocrine interactions that involve the hypothalamic-pituitary-gonadal axis: hypothalamus (gonadotropin releasing hormone), pituitary (FSH, LH), and gonads (E2, PG, T), and many other hormones secreted by these organs [46–48]. Information about the effects of VD deficiency and sufficiency on reproductive hormones is limited. In our study, relative to saline-injected hens, injection with LPS significantly reduced the level of E2, LH, T, and PG in the serum and significantly increased serum CORT concentration, whereas there were no significant effects on PTH and FSH. The elevated CORT and reduced sex steroid hormones in laying hens could be attributed to acute stress caused by LPS. However, VD deficiency results in a significant reduction in FSH and LH levels, and a remarkable increase in E2 and PG regardless of LPS challenge. However, the changes induced by VD₃ deficiency were completely reversed by VD₃ supplementation. Furthermore, LH concentration was significantly increased in LPS-challenged and VD₃-supplemented groups compared to the LPS-challenged and VD₃-deficient treatment. Increased LH concentration and decreased E2 and

Table 8 Effect of vitamin D₃ and LPS challenge on serum cytokines of laying hens infected with LPS*

Items	Dietary VD ₃ levels, IU/kg				P-values			
	0		3000		SEM	VD ₃	LPS	Interaction
	Saline	LPS	Saline	LPS				
TNF-α (pg/ml)	134.29	183.78	114.62	168.08	7.53	0.114	< 0.001	0.854
IFN-γ (pg/ml)	66.71	77.66	60.85	67.70	3.09	0.211	0.162	0.741
IL-1β (pg/ml)	15.01	18.65	13.15	17.26	0.60	0.085	< 0.001	0.792
IL-2 (pg/ml)	0.90	1.37	1.05	1.62	0.09	0.154	< 0.001	0.726
IL-4 (pg/ml)	25.95	49.70	24.50	48.36	3.39	0.777	< 0.001	0.992
IL-6 (pg/ml)	90.31	145.60	81.79	124.79	6.47	0.031	< 0.001	0.337
IL-8 (pg/ml)	28.49	48.34	25.96	44.06	2.74	0.400	< 0.001	0.828
IL-10 (pg/ml)	53.71	43.01	37.21	35.35	2.16	0.001	0.066	0.186
IFN-γ/IL-4	2.57	1.56	2.48	1.40	0.26	0.001	0.686	0.729

*Data are presented as mean ± SEM (n = 6 hens /group)

^{a-c} Data in the same row with no common superscript differ significantly (P < 0.05)

SEM standard error of the mean; LPS lipopolysaccharide; TNF-α tumor necrosis factor; IL interleukin; IFN interferon

Table 9 Gene expression in the spleens of laying hens fed diets with or without supplemental dietary VD₃*

Items	Dietary VD ₃ levels, IU/kg				P-values			
	0 IU/kg		3000 IU/kg		SEM	VD ₃	LPS	Interaction
	Saline	LPS	Saline	LPS				
VDR	1.52	1.22	1.16	1.71	0.14	0.998	0.629	0.200
RXR-α	1.96	1.78	1.14	1.05	0.14	0.020	0.824	0.344
TLR2	1.82 ^a	0.76 ^b	0.98 ^b	1.04 ^b	0.14	0.143	0.094	0.042
TLR4	1.13	3.17	1.22	1.97	0.24	0.300	0.020	0.620
NF-κB	1.56 ^b	2.64 ^a	1.08 ^b	1.58 ^b	0.18	0.009	0.011	0.029
TNF-α	0.98	1.25	1.12	1.74	0.11	0.130	0.061	0.815
IFN-γ	0.73	0.89	0.98	0.98	0.11	0.559	0.053	0.443
IL-1β	1.08	0.88	1.28	0.86	0.19	0.700	0.220	0.960
IL-6	0.51	2.30	0.81	1.94	0.23	0.893	< 0.001	0.439
IL-8	1.58	1.80	1.11	1.97	0.19	0.879	0.940	0.181
LYZ	1.84 ^a	0.52 ^b	0.47 ^b	1.43 ^a	0.20	0.561	0.453	0.003
iNOS	2.10	1.82	1.30	1.12	0.17	0.016	0.750	0.258
pIgR	3.75 ^a	1.42 ^b	0.96 ^b	2.08 ^a	0.33	0.048	0.241	0.003

*Data are presented as mean ± SEM (n = 6 hens /group)

^{a,c} Mean values with different letters differ significantly (p < 0.05)

SEM standard error of the mean; LPS lipopolysaccharide; VDR vitamin D receptor; RXR retinoid X receptor; TLR Toll-like receptor; NF-κB Nuclear factor κB; TNF-α tumor necrosis factor; IFN interferon; IL interleukin; LYZ lysozyme; iNOS inducible nitric oxide synthase; pIgR polyglobulin receptor

PG levels in VD₃-supplemented groups implied that addition of sufficient VD₃ could counteract the negative feedback effect of E2 on hypothalamic and pituitary hormone secretion, and positively affect follicle development and ovulation by inhibiting E2 levels. Significant reduced egg production observed in VD₃-deficient hens might be involved in the lower LH and higher levels of E2 and PG in serum. This finding accounts for the beneficial effects of VD supplementation on hens' laying performance, possibly related to reproductive hormones.

Calcium homeostasis reflects the balance between fluxes to and from the gut, regulated renal reabsorption, and deposition/mobilization by bone. This process is primarily regulated by the complex interactions of PTH, CT, and vitamin D, as well as with direct exchange with the bone matrix [49]. PTH secreted by the chief cells of the parathyroid glands regulates renal synthesis of 1,25-(OH)₂-D₃, increasing the concentration of calcium in the blood. PTH secretion is stimulated by hypocalcemia; its concentration provides an important index of vitamin D/calcium status, whereas CT decreases calcium concentration [50]. In our study, relative to saline-injected hens, injection with LPS significantly reduced serum CT, whereas it had no significant effect on PTH. This finding showed that LPS challenge could increase serum calcium levels by inhibiting CT secretion. Supplementation of VD₃ resulted in reduced serum CT levels of the birds relative

to VD₃-deficient treatment, which was not in agreement with the finding of Jiang et al. [42]. This change likely reflects increases in serum calcium. However, lower serum PTH content was observed in the VD-supplemented hens, which is consistent with the results of Jiang et al. [42] this change might be helpful to suppress the reabsorption, and deposition and mobilization by bone and help to maintain blood calcium homeostasis in the VD₃-supplemented diets with normal level of dietary calcium and phosphorus. Increased PTH, however, was observed in VD₃-deficient hens, resulting in calcium resorption in the kidneys or intestines; this might causes change in serum calcium. Accordingly, these results suggested that reduced egg-shell quality in the VD₃-deficient group was because of effects on PTH and CT concentrations affecting serum calcium levels.

MPO is most abundantly expressed in neutrophil granulocytes; it participates in innate immune defense and plays a key role in the resolution of inflammation [51]. The complement system has been long recognized as a central part of innate immunity in defense against pathogen invasion [52, 53]. In our study, VD₃ deficiency significantly increased MPO activity, while LPS-injected hens fed with VD₃-supplemented diet showed a significant reduction in MPO activity compared with saline-injected hens without VD₃ supplementation. These results showed that VD₃ deficiency could cause inflammation; however, VD₃ supplementation not only reversed VD₃ deficiency-induced inflammatory responses but also alleviated immunological inflammation caused by LPS through reducing MPO activity. In addition, we also observed that serum complement C3, C4, and C5 activity were not affected by VD₃ supplementation, which is in agreement with the report of Zhang et al. [54] who found that dietary VD₃ had no effect on serum complement level in fish. However, the higher level of 25(OH)D in human serum will directly accompany increasing C4 concentration but decreasing C3 concentration [55]. The inconsistent results might contribute to different biological species. Our findings indicated that VD₃ apparently did not activate the complement system.

Natural antibodies are essential components of the innate immune system, and they are present in the body without known antigenic stimulation of B cells [56]. In our study, VD₃ deficiency promoted serum IgG production, whereas VD₃ supplementation significantly reversed this change. This is consistent with other findings [57] indicating VD₃ downregulated T cell-driven IgG production or inhibited antibody production through preventing the proliferation of activated B cells [24, 58, 59]. In hens not injected with LPS, however, VD₃ supplementation was associated with a significant increase in serum IgM, whereas lower levels of total IgM were observed in hens injected with LPS and fed with sufficient VD₃. This observation suggested that VD₃

supplementation could be beneficial in enhancing the innate humoral immunity of hens reared in an unchallenged environment, while counteracting the inflammatory response when confronted by an inappropriate and overly exuberant immune reaction caused by LPS through inhibiting IgM production [60]. Therefore, VD₃ may be a useful immune regulator in layer production.

In our study, VD₃ deficiency resulted in a significant increase in the proportion of CD3⁺ T lymphocytes, as well as in a remarkable reduction in the regulatory CD4⁺CD25⁺ T cell population, while these changes were reversed by VD₃-sufficient supplementation regardless of LPS administration. Similar results have been reported in humans [61, 62], mice [63], and chickens [24]. Vitamin D has been reported to inhibit Th1 and Th17 responses, induce regulatory T cell responses, and control proliferation and helper T cell localization [64, 65]. Regulatory T cells are the subset of CD4⁺ T cells that express FOXP3 protein and the cell surface marker CD25 with anti-inflammatory and immunosuppressive properties by the secretion of potentially inhibitory cytokines IL-10 and TGF-β [66]. Therefore, the results showed that VD₃ deficiency could activate a T cell-mediated immune response and possibly lead to an inflammatory response with elevated levels of CD3⁺ T cells, whereas VD₃ supplementation could suppress an inflammatory response by activating regulatory T cells [67] and converting naïve T cells into regulatory T cells and preserve immune homeostasis [9, 13, 68]. In addition, in the absence of LPS challenge, the percentage of CD8⁺ T cells in circulation was not influenced by VD₃ treatment, whereas in the presence of LPS challenge, LPS induced an increase in the numbers of CD8⁺ T cells. This increase was reversed by supplemental VD₃. In contrast to our findings, Morris et al. [24] indicated that feeding 25(OH) D increased CD8⁺ cell percentage in the cecal tonsils of pigs. CD8⁺ T cells, the cytotoxic T cells, are effector T lymphocytes, essential in immune protection against intracellular pathogens. Lower CD8⁺ T cell populations observed in VD₃-sufficient animals has been suggested to have a protective role against LPS-induced immunological stress. Selevraj et al. [69] found that VD₃ may inhibit T cell proliferation and the recruitment and activation of T-cells through CXC chemokines at the site of infection and may act as a potential anti-inflammatory agent. Therefore, our results suggested that VD₃ supplementation suppressed LPS-induced inflammatory response by modulating T cell subset differentiation and T lymphocyte effector function. We speculated this role of VD₃ is possibly carried out through regulating VDR or stimulating VD-related enzyme expression.

The above results implied that vitamin D deficiency depressed the immune function of chickens. In addition, immunity is also associated with inflammation, which is

primarily mediated by inflammatory cytokines in chicken. We further examined the effects of VD₃ deficiency and sufficiency on serum cytokines and splenic immune-related gene expression in laying hens challenged with *E. coli* LPS. In our study, LPS stimulation upregulated all proinflammatory cytokines (TNF-α, IL-1β, IL-2, IL-4, IL-6, IL-8) in the serum of the laying hens. Vitamin D administration differentially influenced cytokine profiles, with a reduced production of proinflammatory cytokines IL-6, and the ratio of IFN-γ to IL-4 (Th1/Th2). Vitamin D administration showed a contradictory result for Th2 cytokine IL-10 expression, in contrast to the observation of increased IL-10 production in other studies [70]. This finding was in contrast to the vitamin D-deficient group irrespective of LPS. Similar findings relating to VD₃ and its metabolites inducing pro-inflammatory and anti-inflammatory cytokine production stimulated by bacteria and bacterial PRR exposure were reported by previous authors in human [71, 72], murine [31, 73], porcine [24], and avian systems [21–23, 74]. Boodhoo et al. [74] demonstrated that vitamin D reduces chicken T lymphocyte proliferation as well as the number of IFN-γ producing cells. In addition, it has been previously shown that VD₃ can modify the balance between the Th1 and Th2 response [70]. The decreased ratio of IFN-γ to IL-4 in the blood of VD-sufficient hens, regardless of LPS injection, may suggest that VD suppresses production of Th1 cytokines and promotes Th2 responses. Furthermore, in clinical settings, a reduction in IFN-γ production may reduce the immunopathology observed in acute or chronic inflammatory diseases induced by infectious agents. Therefore, these results indicated that VD₃ deficiency is associated with immunological hyperactivity and could cause an inflammatory response. The response could result from enhancing Th1-derived responses and suppressing Th2-driven responses, while VD supplementation dampened the inflammatory response from LPS through suppression of pro-inflammatory Th1 in favor of an anti-inflammatory Th2 phenotype, but without causing immunopathology.

The spleen is an important peripheral immune organ, comprising B cells, T cells, macrophages, and dendritic cells. Splenic cells secrete cytokines following immune stimulation and play an important role in systemic immune function. Toll-like receptor 4 recognizes the LPS of gram-negative bacteria such as *E. coli* and *Salmonella* and stimulates the intracellular cascades to activate nuclear factor-κB. In turn, this stimulation leads to the synthesis of cytokines and other molecules and contributes to the initiation of an inflammatory response. In our study, we observed upregulated expression of splenic TLR4, NF-κB and IL-6 gene mRNA levels in the spleen following LPS challenge. This finding may imply a

robust inflammatory response caused by LPS. Meanwhile, significant upregulated expression of RXR- α , LYZ, iNOS, and pIgR genes in the spleen were found in VD₃-deficient hens, indicating that VD₃ deficiency could cause inflammation in laying hens. This observation was in accordance with the results reported by previous researchers [75]. Interestingly, in the presence of LPS, VD₃ supplementation significantly upregulated splenic TLR2, LYZ, and pIgR gene mRNA but downregulated the expression of the NF- κ B gene in the spleen relative to single LPS-injected hens. These results suggested that VD₃ alleviates LPS-induced splenic immunological stress through suppressing the transcription of NF- κ B genes and enhancing innate immunity. Previous studies have demonstrated that vitamin D exerts its anti-inflammatory activity response to LPS stimulation through decreasing gene expression associated with inflammation [22, 71] as well as by blocking NF- κ B activation [31, 71, 76, 77]. Thus, the observed effects of vitamin D on the inhibition of LPS-induced NF- κ B activation in hens might be because of the anti-inflammatory effect of VD₃ [78–80]. Although previous studies having demonstrated that LPS downregulates VDR expression in keratinocytes [81], vitamin D and its metabolites could alter the expression of different genes for immunoregulation by binding to VDR [82, 83], VDR gene expression was not influenced by LPS or VD₃ treatment in our study and needs to be investigated further.

Conclusion

Our study showed that VD₃ deficiency caused decreased egg production, poor eggshell quality, and inflammatory responses in laying hens. Dietary VD₃-sufficient supplementation could improve egg production and egg quality. Supplementation could also repress immune inflammatory responses caused by LPS administration or VD₃ deficiency by affecting reproductive hormone secretion and regulating the NF- κ B signaling pathway. Thus, VD₃ supplementation could be beneficial to protect layer hens in both preventing immunological stress and VD₃ deficiency. We plan to further investigate the potential mechanisms of action related to the anti-inflammatory effect of VD₃ in chickens.

Abbreviations

1,25-(OH)₂D₃: 1 α ,25-dihydroxycholecalciferol; 25(OH)D: 25-hydroxycholecalciferol; C3: Complement 3; C4: Complement 4; C5: Complement 5; CORT: Cortisone; CT: Calcitonin; E2: Estradiol; FCR: Feed conversion rate; FSH: Follicle stimulating hormone; IFN: Interferon; IgG: Immunoglobulins G; IgM: Immunoglobulins M; IL: Interleukin; iNOS: Inducible nitric oxide synthase; LH: Luteinizing hormone; LPS: Lipopolysaccharide; LYZ: Lysozyme; MPO: Myeloperoxidase; NF- κ B: Nuclear factor- κ B; PG: Progesterone; pIgR: Polyglobulin receptor; PTH: Parathyroid hormone; RXR: Retinoid X receptor; T: Testosterone; TLR: Toll-like receptor; TNF- α : Tumor necrosis factor; VD₃: Vitamin D₃; VDR: Vitamin D receptor

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Availability of data and materials

All data generated or analyzed during the current study are available from the corresponding author on reasonable request. The datasets supporting the conclusions of this article are included within the article.

Authors' contributions

The authors' contributions are as follows: Zhong Wang, conceived and designed the research; Yanqiang Geng, conducted the animal trial and sampling, performed sample analyses, analyzed the data, and drafted the manuscript; Zhong Wang and Yuming Guo participated in the revision of the manuscript. All authors contributed to the data interpretation and approved the final version of the manuscript. We thank Andrea Baird, MD, from Liwen Bianji, Edanz Editing China (<https://www.liwenbianji.cn>), for editing the English text of a draft of this manuscript.

Ethics approval and consent to participate

All animal experiments were reviewed and approved by the Institutional Animal Care and Use Committee of China Agricultural University and performed in accordance with the "Guidelines for Experimental Animals" of the Ministry of Science and Technology (Beijing, PR China).

Consent for publication

Not applicable.

Competing interests

All authors approve the submission of this manuscript and declare no conflict of interest. The manuscript has not been published previously, and not under consideration for publication elsewhere. Funders had no role in the study design, analysis, or writing of this article.

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References

- Zhang R, Naughton DP. Vitamin D in health and disease: current perspectives. *Nutr J.* 2010;9:65.
- Norman AW. From vitamin D to hormone D: fundamentals of the vitamin D endocrine system essential for good health. *Am J Clin Nutr.* 2008;88:491S–95S.
- Norman AW. Minireview: vitamin D receptor: new assignments for an already busy receptor. *Endocrinology.* 2006;147:5542–8.
- Wang TT, Tavera-Mendoza LE, Laperriere D, Libby E, MacLeod NB, Nagai Y, Bourdeau V, Konstorum A, Lallemand B, Zhang R, et al. Large-scale in silico and microarray-based identification of direct 1,25-dihydroxyvitamin D₃ target genes. *Mol Endocrinol.* 2005;19:2685–95.
- Overbergh L, Decallonne B, Valckx D, Verstuyf A, Depovere J, Laureys J, Rutgeerts O, Saint-Arnaud R, Bouillon R, Mathieu C. Identification and immune regulation of 25-hydroxyvitamin D-1- α -hydroxylase in murine macrophages. *Clin Exp Immunol.* 2000;120:139–46.
- Lips P. Vitamin D physiology. *Prog Biophys Mol Biol.* 2006;92:4–8.
- Gorman S, Buckley AG, Ling KM, Berry LJ, Fear VS, Stick SM, Larcombe AN, Kicic A, Hart PH. Vitamin D supplementation of initially vitamin D-deficient

- mice diminishes lung inflammation with limited effects on pulmonary epithelial integrity. *Physiological Reports*. 2017;5(15):e13371.
8. Suaini NH, Zhang Y, Vuillemin PJ, Allen KJ, Harrison LC. Immune modulation by vitamin D and its relevance to food allergy. *Nutrients*. 2015;7:6088–108.
 9. Adams JS, Hewison M. Unexpected actions of vitamin D: new perspectives on the regulation of innate and adaptive immunity. *Nat Clin Pract Endocrinol Metab*. 2008;4:80–90.
 10. Li YC, Chen YZ, Liu WC, Thadhani R. MicroRNA-mediated mechanism of vitamin D regulation of innate immune response. *J Steroid Biochem Mol Biol*. 2014;144PA:81–6.
 11. Krishnan AV, Feldman D. Mechanisms of the anti-cancer and anti-inflammatory actions of vitamin D. *Annu Rev Pharmacol Toxicol*. 2011;51:311–36.
 12. Xu S, Chen YH, Tan ZX, Xie DD, Zhang C, Xia MZ, Wang H, Zhao H, Xu DX, Yu DX. Vitamin D3 pretreatment alleviates renal oxidative stress in lipopolysaccharide-induced acute kidney injury. *J Steroid Biochem Mol Biol*. 2015;152:133–41.
 13. Di Rosa M, Malaguarnera M, Nicoletti F, Malaguarnera L. Vitamin D3: a helpful immuno-modulator. *Immunology*. 2011;134:123–39.
 14. Greiller CL, Martineau AR. Modulation of the immune response to respiratory viruses by vitamin D. *Nutrients*. 2015;7:4240–70.
 15. Urashima M, Segawa T, Okazaki M, Kurihara M, Wada Y, Ida H. Randomized trial of vitamin D supplementation to prevent seasonal influenza a in schoolchildren. *Am J Clin Nutr*. 2010;91:1255–60.
 16. Sarkar S, Hewison M, Studzinski GP, Li YC, Kalia V. Role of vitamin D in cytotoxic T lymphocyte immunity to pathogens and cancer. *Crit Rev Clin Lab Sci*. 2016;53:132–45.
 17. Peterson CA, Heffernan ME. Serum tumor necrosis factor- α concentrations are negatively correlated with serum 25(OH)D concentrations in healthy women. *J Inflamm (Lond)*. 2008;5:10.
 18. Wang TT, Nestel FP, Bourdeau V, Nagai Y, Wang Q, Liao J, Tavera-Mendoza L, Lin R, Hanrahan JW, Mader S, White JH. Cutting edge: 1,25-dihydroxyvitamin D3 is a direct inducer of antimicrobial peptide gene expression. *J Immunol*. 2004;173:2909–12.
 19. White JH. Vitamin D as an inducer of cathelicidin antimicrobial peptide expression: past, present and future. *J Steroid Biochem Mol Biol*. 2010;121:234–8.
 20. Zhao Y, Yu B, Mao X, He J, Huang Z, Zheng P, Yu J, Han G, Liang X, Chen D. Dietary vitamin D supplementation attenuates immune responses of pigs challenged with rotavirus potentially through the retinoic acid-inducible gene 1 signalling pathway. *Br J Nutr*. 2014;112:381–9.
 21. Lu L, Li SM, Zhang L, Liu XQ, Li DY, Zhao XL, Liu YP. Expression of beta-defensins in intestines of chickens injected with vitamin D3 and lipopolysaccharide. *Genet Mol Res*. 2015;14:3330–7.
 22. Morris A, Shanmugasundaram R, Lilburn MS, Selvaraj RK. 25-hydroxycholecalciferol supplementation improves growth performance and decreases inflammation during an experimental lipopolysaccharide injection. *Poult Sci*. 2014;93:1951–6.
 23. Shojadoost B, Behboudi S, Villanueva AI, Brisbin JT, Ashkar AA, Sharif S. Vitamin D3 modulates the function of chicken macrophages. *Res Vet Sci*. 2015;100:45–51.
 24. Morris A, Shanmugasundaram R, McDonald J, Selvaraj RK. Effect of in vitro and in vivo 25-hydroxyvitamin D treatment on macrophages, T cells, and layer chickens during a coccidia challenge. *J Anim Sci*. 2015;93:2894–903.
 25. Zhang L, Lu L, Li S, Zhang G, Ouyang L, Robinson K, Tang Y, Zhu Q, Li D, Hu Y, Liu Y. 1,25-Dihydroxyvitamin-D3 induces avian beta-Defensin gene expression in chickens. *PLoS One*. 2016;11:e0154546.
 26. Rodriguez-Lecompte JC, Yitbarek A, Cuperus T, Echeverry H, van Dijk A. The immunomodulatory effect of vitamin D in chickens is dose-dependent and influenced by calcium and phosphorus levels. *Poult Sci*. 2016;95:2547–56.
 27. Aslam SM, Garlich JD, Qureshi MA. Vitamin D deficiency alters the immune responses of broiler chicks. *Poult Sci*. 1998;77:842–9.
 28. Barreda DR, Konowalchuk JD, Rieger AM, Wong ME, Havixbeck JJ. Triennial growth symposium—novel roles for vitamin D in animal immunity and health. *J Anim Sci*. 2014;92:930–8.
 29. Ploegaert TC, De Vries RG, Nieuwland MG, Lammers A, Savelkoul HF, Parmentier HK. Intratracheally administered pathogen-associated molecular patterns affect antibody responses of poultry. *Poult Sci*. 2007;86:1667–76.
 30. Star L, Nieuwland MG, Kemp B, Parmentier HK. Effect of single or combined climatic and hygienic stress on natural and specific humoral immune competence in four layer lines. *Poult Sci*. 2007;86:1894–903.
 31. Tan ZX, Chen YH, Xu S, Qin HY, Wang H, Zhang C, Xu DX, Zhao H. Calcitriol inhibits tumor necrosis factor alpha and macrophage inflammatory protein-2 during lipopolysaccharide-induced acute lung injury in mice. *Steroids*. 2016; 112:81–7.
 32. Jarosz L, Marek A, Gradzki Z, Kwiecien M, Kalinowski M. The effect of feed supplementation with zinc chelate and zinc sulphate on selected humoral and cell-mediated immune parameters and cytokine concentration in broiler chickens. *Res Vet Sci*. 2017;112:59–65.
 33. Vijayendra Chary A, Hemalatha R, Seshacharyulu M, Vasudeva Murali M, Jayaprakash D, Dinesh Kumar B. Vitamin D deficiency in pregnant women impairs regulatory T cell function. *J Steroid Biochem Mol Biol*. 2015;147:48–55.
 34. Frost TJ, Roland DA Sr, Untawale GG. Influence of vitamin D3, 1 alpha-hydroxyvitamin D3, and 1,25-dihydroxyvitamin D3 on eggshell quality, tibia strength, and various production parameters in commercial laying hens. *Poult Sci*. 1990;69:2008–16.
 35. Mattila P, Valaja J, Rossow L, Venalainen E, Tupasela T. Effect of vitamin D2- and D3-enriched diets on egg vitamin D content, production, and bird condition during an entire production period. *Poult Sci*. 2004;83:433–40.
 36. Sun ZW, Yan L, YY G, Zhao JP, Lin H, Guo YM. Increasing dietary vitamin D3 improves the walking ability and welfare status of broiler chickens reared at high stocking densities. *Poult Sci*. 2013;92:3071–9.
 37. Saunders-Blades JL, Korver DR. Effect of hen age and maternal vitamin D source on performance, hatchability, bone mineral density, and progeny in vitro early innate immune function. *Poult Sci*. 2015;94:1233–46.
 38. Persia ME, Higgins M, Wang T, Trample D, Bobeck EA. Effects of long-term supplementation of laying hens with high concentrations of cholecalciferol on performance and egg quality. *Poult Sci*. 2013;92:2930–7.
 39. Zerwekh JE. Blood biomarkers of vitamin D status. *Am J Clin Nutr*. 2008;87: 10875–91S.
 40. Ryz NR, Lochner A, Bhullar K, Ma C, Huang T, Bhinder G, Bosman E, Wu X, Innis SM, Jacobson K, Vallance BA. Dietary vitamin D3 deficiency alters intestinal mucosal defense and increases susceptibility to *Citrobacter rodentium*-induced colitis. *Am J Physiol Gastrointest Liver Physiol*. 2015;309:G730–42.
 41. Whitehead CC, McCormack HA, McTeir L, Fleming RH. High vitamin D3 requirements in broilers for bone quality and prevention of tibial dyschondroplasia and interactions with dietary calcium, available phosphorus and vitamin A. *Br Poult Sci*. 2004;45:425–36.
 42. Jiang S, Jiang Z, Yang K, Chen F, Zheng C, Wang L. Dietary vitamin D3 requirement of Chinese yellow-feathered broilers. *Poult Sci*. 2015;94:2210–20.
 43. Khan SH, Shahid R, Mian AA, Sardar R, Anjum MA. Effect of the level of cholecalciferol supplementation of broiler diets on the performance and tibial dyschondroplasia. *J Anim Physiol Anim Nutr (Berl)*. 2010;94:584–93.
 44. Fleming RH. Nutritional factors affecting poultry bone health. *Proc Nutr Soc*. 2008;67:177–83.
 45. Zhou TX, Chen YJ, Yoo JS, Huang Y, Lee JH, Jang HD, Shin SO, Kim HJ, Cho JH, Kim IH. Effects of chitooligosaccharide supplementation on performance, blood characteristics, relative organ weight, and meat quality in broiler chickens. *Poult Sci*. 2009;88:593–600.
 46. Rangel PL, Gutierrez CG. Reproduction in hens: is testosterone necessary for the ovulatory process? *Gen Comp Endocrinol*. 2014;203:250–61.
 47. Segner H, Verburg-van Kemenade BML, Chadzinska M. The immunomodulatory role of the hypothalamus-pituitary-gonad axis: proximate mechanism for reproduction-immune trade offs? *Dev Comp Immunol*. 2017;66:43–60.
 48. Tarumi W, Itoh MT, Suzuki N. Effects of 5 α -dihydrotestosterone and 17 β -estradiol on the mouse ovarian follicle development and oocyte maturation. *PLoS One*. 2014;9:e99423.
 49. Khanal RC, Nemere I. Endocrine regulation of calcium transport in epithelia. *Clin Exp Pharmacol Physiol*. 2008;35:1277–87.
 50. Potts JT. Parathyroid hormone: past and present. *J Endocrinol*. 2005;187:311–25.
 51. Klebanoff SJ. Myeloperoxidase: friend and foe. *J Leukoc Biol*. 2005;77:598–625.
 52. Ward PA, Gao H. Sepsis, complement and the dysregulated inflammatory response. *J Cell Mol Med*. 2009;13:4154–60.
 53. Parmentier HK, Lammers A, Hoekman JJ, De Vries RG, Zaenen IT, Savelkoul HF. Different levels of natural antibodies in chickens divergently selected for specific antibody responses. *Dev Comp Immunol*. 2004;28:39–49.
 54. Zhang J, Chang JR, Duan XH, Yu YR, Zhang BH. Thyroid hormone attenuates vascular calcification induced by vitamin D3 plus nicotine in rats. *Calcif Tissue Int*. 2015;96:80–7.
 55. Sakem B, Nock C, Stanga Z, Medina P, Nydegger UE, Risch M, Risch L. Serum concentrations of 25-hydroxyvitamin D and immunoglobulins in an older Swiss cohort: results of the senior labor study. *BMC Med*. 2013;11:176.

56. Ochsenbein AF, Zinkernagel RM. Natural antibodies and complement link innate and acquired immunity. *Immunol Today*. 2000;21:624–30.
57. Aranow C. Vitamin D and the immune system. *J Investig Med*. 2011;59:881–6.
58. Yang HF, Zhang ZH, Chang ZQ, Tang KL, Lin DZ, Xu JZ. Vitamin D deficiency affects the immunity against *Mycobacterium tuberculosis* infection in mice. *Clin Exp Med*. 2013;13:265–70.
59. Chen S, Sims GP, Chen XX, Gu YY, Chen S, Lipsky PE. Modulatory effects of 1,25-dihydroxyvitamin D3 on human B cell differentiation. *J Immunol*. 2007;179:1634–47.
60. Leventis P, Patel S. Clinical aspects of vitamin D in the management of rheumatoid arthritis. *Rheumatology (Oxford)*. 2008;47:1617–21.
61. Harishankar M, Anbalagan S, Selvaraj P. Effect of vitamin D3 on chemokine levels and regulatory T-cells in pulmonary tuberculosis. *Int Immunopharmacol*. 2016;34:86–91.
62. Jeffery LE, Burke F, Mura M, Zheng Y, Qureshi OS, Hewison M, Walker LS, Lammas DA, Raza K, Sansom DM. 1,25-Dihydroxyvitamin D3 and IL-2 combine to inhibit T cell production of inflammatory cytokines and promote development of regulatory T cells expressing CTLA-4 and FoxP3. *J Immunol*. 2009;183:5458–67.
63. Gorman S, Kuritzky LA, Judge MA, Dixon KM, McGlade JP, Mason RS, Finlay-Jones JJ, Hart PH. Topically applied 1,25-dihydroxyvitamin D3 enhances the suppressive activity of CD4+CD25+ cells in the draining lymph nodes. *J Immunol*. 2007;179:6273–83.
64. Hayes CE, Hubler SL, Moore JR, Barta LE, Praska CE, Nashold FE. Vitamin D actions on CD4(+) T cells in autoimmune disease. *Front Immunol*. 2015;6:100.
65. Cantorna MT, Snyder L, Lin YD, Yang L. Vitamin D and 1,25(OH)2D regulation of T cells. *Nutrients*. 2015;7:3011–21.
66. Prietl B, Treiber G, Pieber TR, Amrein K. Vitamin D and immune function. *Nutrients*. 2013;5:2502–21.
67. Bruce D, Ooi JH, Yu S, Cantorna MT. Vitamin D and host resistance to infection? Putting the cart in front of the horse. *Exp Biol Med (Maywood)*. 2010;235:921–7.
68. Mangin M, Sinha R, Fincher K. Inflammation and vitamin D: the infection connection. *Inflamm Res*. 2014;63:803–19.
69. Selvaraj P, Harishankar M, Singh B, Banurekha VV, Jawahar MS. Effect of vitamin D3 on chemokine expression in pulmonary tuberculosis. *Cytokine*. 2012;60:212–9.
70. Boonstra A, Barrat FJ, Crain C, Heath VL, Savelkoul HF, O'Garra A. 1 α ,25-Dihydroxyvitamin d3 has a direct effect on naive CD4(+) T cells to enhance the development of Th2 cells. *J Immunol*. 2001;167:4974–80.
71. Fitch N, Becker AB, HayGlass KT. Vitamin D [1,25(OH)2D3] differentially regulates human innate cytokine responses to bacterial versus viral pattern recognition receptor stimuli. *J Immunol*. 2016;196:2965–72.
72. Harishankar M, Afsal K, Banurekha VV, Meenakshi N, Selvaraj P. 1,25-Dihydroxy vitamin D3 downregulates pro-inflammatory cytokine response in pulmonary tuberculosis. *Int Immunopharmacol*. 2014;23:148–52.
73. Zhang Y, Leung DY, Richers BN, Liu Y, Remigio LK, Riches DW, Goleva E. Vitamin D inhibits monocyte/macrophage proinflammatory cytokine production by targeting MAPK phosphatase-1. *J Immunol*. 2012;188:2127–35.
74. Boodhoo N, Sharif S, Behboudi S. 1 α ,25(OH)2 vitamin D3 modulates avian T lymphocyte functions without inducing CTL unresponsiveness. *PLoS One*. 2016;11:e0150134.
75. Adams JS, Liu PT, Chun R, Modlin RL, Hewison M. Vitamin D in defense of the human immune response. *Ann N Y Acad Sci*. 2007;1117:94–105.
76. Chen YH, Yu Z, Fu L, Wang H, Chen X, Zhang C, Lv ZM, Xu DX. Vitamin D3 inhibits lipopolysaccharide-induced placental inflammation through reinforcing interaction between vitamin D receptor and nuclear factor kappa B p65 subunit. *Sci Rep*. 2015;5:10871.
77. Xu S, Chen YH, Tan ZX, Xie DD, Zhang C, Zhang ZH, Wang H, Zhao H, Yu DX, Xu DX. Vitamin D3 pretreatment regulates renal inflammatory responses during lipopolysaccharide-induced acute kidney injury. *Sci Rep*. 2015;5:18687.
78. Kundu R, Theodoraki A, Haas CT, Zhang Y, Chain B, Kriston-Vizi J, Noursadeghi M, Khoo B. Cell-type-specific modulation of innate immune signalling by vitamin D in human mononuclear phagocytes. *Immunology*. 2017;150:55–63.
79. Heulens N, Korf H, Mathyssen C, Everaerts S, De Smidt E, Dooms C, Yserbyt J, Gysemans C, Gayan-Ramirez G, Mathieu C, Janssens W. 1,25-Dihydroxyvitamin D modulates antibacterial and inflammatory response in human cigarette smoke-exposed macrophages. *PLoS One*. 2016;11:e0160482.
80. Oh J, Riek AE, Darwech I, Funai K, Shao J, Chin K, Sierra OL, Carmeliet G, Ostlund RE Jr, Bernal-Mizrachi C. Deletion of macrophage vitamin D receptor promotes insulin resistance and monocyte cholesterol transport to accelerate atherosclerosis in mice. *Cell Rep*. 2015;10:1872–86.
81. Bin Z, Li R, Fang Y, Yu FY, Na X, Fang Z, Ge XJ, Du J. LPS-induced Vitamin D Receptor Decrease in Oral Keratinocytes Is Associated With Oral Lichen Planus. *Scientific Reports*. 2018;8:763.
82. Ge X, Du J, Chen Y, Li YC. vitamin D receptor inhibits nuclear factor kappa B activation by interacting with I kappa B kinase beta protein. *J Biol Chem*. 2013;288:19450–8.
83. Zhao H, Zhang H, Wu H, Li H, Liu L, Guo J, Li C, Shih DQ, Zhang X, Zhao H, Zhang H, Wu H, Li H, Liu L, Guo J, et al. Protective role of 1,25(OH)2 vitamin D3 in the mucosal injury and epithelial barrier disruption in DSS-induced acute colitis in mice. *BMC Gastroenterol*. 2012;12:57.

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